Stereoselective Syntheses of (22R)- and (22S)-22-Methyl-1 α ,25-dihydroxyvitamin D₃: Active Vitamin D₃ Analogs with Restricted Side Chain Conformation

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(22R)- and (22S)-22-methyl-1 α , 25-dihydroxyvitamin D_3 (1b and 1c) were synthesized stereoselectively from 1α -hydroxylated C(22)-steroid 2. The two new vitamin D analogs, which have a side chain with restricted flexibility, were designed to allow the study of the stereochemical structure required to bind to the receptor of the active vitamin D_3 (VDR). According to force-field calculations, the side chain of (22R)- and (22S)-methylated active vitamin D_3 analogs (1b and 1c) adopts with more than 90% of the population gauche(+) and anti conformations, respectively, at the C(17-20-22-23) dihedral angle. Either the (22R)- or (22S)-methylated steroidal side chain was constructed with high stereoselectivity via a kinetically controlled conjugate addition of methylcopper reagent to (22E)or (22Z)-22-en-24-ones (6 or 7), respectively, as a key step. The ability of the two analogs to bind to VDR was examined and only the (22S)-isomer (1c) showed significant activity. From the results, the side chain conformation best fitted to VDR was suggested to be the anti with respect to the C(17-20-22-23) dihedral angle.

Structure and function studies of the active vitamin D analogs are important not only to develop a clinically useful medicine but also to clarify the mechanism of action of the active vitamin D_3 , 1α , 25-dihydroxyvitamin D_3 (1,25- $(OH)_2D_3$, 1a). The active vitamin D_3 is a multifunctional



hormone¹ which is believed to exert its activities by a mechanism mediated by the nuclear vitamin D receptor (VDR).² Extensive studies on the structure and function of vitamin D analogs have shown that the separation of the multiple activities of the active vitamin is possible.³ However the mechanism of the separation of the actions

is unclear. In the case of 22-oxa- 1α , 25-dihydroxyvitamin D_3 , the separation of the differentiating action from calcemic activities can be explained by a pharmacokinetics.⁴ But in the case of 24-di- and trihomo-1.25-dihydroxyvitamin D₃ analogs, the separation of the two actions in the intestine, calcium transport, and synthesis of calcium binding protein (CaBP), cannot be explained by the same mechanism.⁵ In this case, no calcium transporting activity was elicited even when these vitamin D analogs reached the target tissue, and the synthesis of CaBP was proved at the mRNA level. In any case it should be noted that these analogs played an important role in understanding the mechanism of the expression of biological activities.

In contrast to general steroid compounds, vitamin D has a flexible structure. The A ring with two exocyclic double bonds, the opened B ring, and the side chain can adopt a wide range of conformations. So it is interesting to synthesize vitamin D analogs with a fixed conformation and to test their activities to know the essential structure required for expressing each of the activities. The C(17-20-22-23) torsion angle of the side chain is the key to

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 Table I. Minimum Energy Conformations of the Side Chain of 1a,25-Dihydroxyvitamin D3 and Its (22R)- and (22S)-Methyl

 Analogs^a

compound	conformation at C(17-20-22-23)					
	anti		gauche(+)		gauche(-)	
	energy ^b (population, ^d %)	angle ^c	energy ^b (population, $d \%$)	angle ^c	energy ^b (population, ^d $\%$)	angle ^c
1'a 1'b (22R)	33.005 (38)	-168°	32.723 (62) 35.732 (95)	64° 68°	37.454 (5)	-73°
1'c (22S)	36.076 (90)	-167°	37.403 (10)	55°		

^a The energies were calculated for model system (1', Figure 1) by MM2PP force-field method.⁸ ^b Steric energy in kcal/mol. ^c C(17-20-22-23) torsion angle. ^d Boltzmann distribution at 25 °C.



Figure 1.

determine the conformation of the side chain. It is known from the X-ray analysis that vitamin D₃ adopts two major side chain conformations, anti and gauche(+) conformations with respect to C(17-20-22-23) torsion angle (Figure 1).⁶ But in the X-ray structure of 25-hydroxyvitamin D_3 , only the anti conformer was observed.7 This is in contrast to the results of force-field calculations. By MM2PP calculations,⁸ the 25-hydroxyvitamin D_3 side chain has two minimum energy conformations with respect to the torsion angle at C(17-20-22-23): the gauche(+) and the anti conformations, the former being 62:38 more favorable than the latter at 25 °C (Table I). In any case these two conformations are similar in energy. Therefore it is interesting to know which is the appropriate conformation to bind to VDR. According to force-field calculations, introduction of a methyl group to the 22-position makes one of the staggered conformations exceedingly favorable (Table I). Thus (22R)- and (22S)-22-methyl-1,25-dihydroxyvitamin D_3 (1b and 1c) adopt gauche(+) and anti forms, respectively, with more than 90% of the population. Introduction of a methyl group was expected to cause minimum effect on the electronic and geometric structure of the parent vitamin D molecule. On the same idea Ikekawa's group have already synthesized two epimeric 22-methoxy derivatives of 1,25-(OH)₂D₃ (1d and 1e) and their biological activity was tested.⁹ From the results, they concluded that the anti conformation is responsible for the activities. However because of the ability of oxygen to form a hydrogen bond with VDR, introduction of methoxyl group was considered not to be appropriate for the purpose of the study. Here we report stereoselective syntheses of (22R)- and (22S)-22-methyl-1 α ,25-dihydroxyvitamin D_3 (1b and 1c) and the competitive binding study of these analogs to intestinal VDR.

Results and Discussion

Strategies for the syntheses of two epimeric 22-methyl active vitamin D₃ analogs (1b and 1c) are as follows (Scheme I). The 22-methyl group was planned to be introduced by conjugate addition of methylcopper reagent to Δ^{22} -24-ketone (6 or 7). Because of the bulky steroid skeleton at the γ -position, the conjugate addition reaction was expected to be highly stereospecific. In fact, as described below, when the reaction was conducted under kinetically controlled conditions, 22*R*- and 22*S*-methylated products (9 and 11) were obtained with high selectivity from (22*E*)- and (22*Z*)-enones (6 and 7), respectively. The enone 6 can be synthesized easily as reported¹⁰ and the (*Z*)-enone 7 is obtainable by photochemical isomerization of the (*E*)-enone (6).

The syntheses were started with readily available 1α hydroxylated C(22) steroid 2.¹¹ Parikh oxidation of C(22)alcohol 2 provided aldehyde 3. The (E)-enone 6 was obtained (73%) from the aldehyde 3 by a modification of the reported procedure.¹⁰ Thus aldol condensation of 3 with 3-hydroxy-3-methylbutanone THP ether (LDA, THF) produced adduct 4 as an isomeric mixture at C(22)which was dehydrated (TsOH) to (E)-enone 5 with concomitant deprotection of the THP group. The 25hydroxyl group was protected (MOMCl, N,N-diisopropylethylamine) to give enone 6. The reaction of the (E)enone 6 with Me₂CuLi under the condition of a normal Gilman reaction (0 °C, THF) gave the 22-methylated compound 9 with high stereoselectivity (98:2) together with a trace of its 22-epimer 11 in high yield (91%). The conjugate addition reaction conducted in the presence of TMSCl¹² (Me₂CuLi. TMSCl, HMPA, THF, -78 °C),¹³ which is supposed to proceed under kinetic control,^{12a} gave a single 22-methylated enol silyl ether 8 (100% selectivity, 95% yield). Desilylation of 8 gave the same 22-methyl ketone 9 (94%) as that obtained by the above Gilman reaction. The change of the conditions did not switch the selectivity but just improved it. The stereochemistry at C(22) of 9 was determined to be R by the X-ray crystal-

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lographic analysis of its derivative 12a, which is one of the epimers of the NaBH₄ reduction products of 9. The PLUTO drawing of the X-ray structure of 12a is shown

in Figure 2.¹⁴ As shown in the Figure 2 the side chain

(14) Crystal data: $C_{34}H_{54}O_{9}$, FW = 606.80, space group P_{21} (monoclinic), Z = 2, a = 20.346(2), b = 6.368(1), c = 13.1256(8) Å, $\beta = 92.685(6)^{\circ}$, V = 1698.6(3) Å³, $D_c = 1.186$ g cm⁻³. The authors have deposited atomic coordinates for this structure with the Cambridge Crystallographic Data

Centre. The coordinates can be obtained, on request, from the Director, Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge, CB2 1EZ, UK. carrying (22*R*)-methyl group has gauche(+) conformation at C(17-20-22-23) in accord with the force-field calculation (Table I).

Since the conjugate addition to (E)-enone 6 was found to give (22R)-methyl epimer 9 nearly exclusively regardless of the conditions, we next examined the addition reaction to (Z)-enone 7. (Z)-Enone 7 was obtained from the (E)enone 6 by dye-sensitized photoisomerization. The proportion of (Z)-enone 7 to (E)-enone 6 at the photostationary state was maximum with a sensitizer of a triplet energy



Figure 2. X-ray crystal structure of compound 12a.

 $(E_{\rm T})$ of 50–60 kcal/mol. However the ratio (Z/E) did not exceed 1. Also the photochemical reaction produced a byproduct 22 oxygenated at C(20) as a mixture of C(20)epimers. The C(20)-alcohol 22 might be produced by the reaction of trace oxygen with $\Delta^{20(22),23}$ -dienol which was formed by photochemical enolization of (Z)-enone 7.¹⁵ In the preparative reaction, isomerization with naphthalene $(E_{\rm T} 60.9 \text{ kcal/mol})$ as a sensitizer (high-pressure mercury lamp, Pyrex filter, benzene) gave a best result (68% yield at 50% conversion). Gilman reaction of the (Z)-enone 7 (Me₂CuLi, THF, 0 °C) gave the 22-methylated ketones 9 and 11 with the same stereoselectivity (98:2) as in the reaction with the (E)-enone (6). The results indicate that

product (97% selectivity) together with 8 (3% selectivity) in 75% yield.¹⁶ Deprotection of 10 gave isomeric 22-methyl ketone 11 (92%) to which we assigned the 22S configuration. Thus highly stereoselective methods for introducing the methyl group to the steroidal 22-position were established.

The cuprate additions with reversed stereoselectivity provided not only stereoselective methods for synthesizing (22R)- and (22S)-alkyl steroids but also intriguing evidence for the mechanism of the cuprate addition reactions.^{12a,17} Here we discuss briefly a mechanism suggested for the cuprate additions (Scheme II).¹⁸ Cuprate addition under normal conditions appears to be thermodynamically controlled, since both (E)- and (Z)-enones (6 and 7) gave the same products (9 and 11) in the same ratio (98:2). In the case of the (E)-enone, the whole steps up to the Cu-(III)- β -adducts (D) are supposed to be in rapid equilibrium

⁽¹⁶⁾ Under these conditions, a byproduct 23 where the 25-OMOM group

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^{(18) (}a) Studies of the cuprate additions and a proposed mechanism for the reactions have been reported.13 (b) Full account of the detailed studies will be reported elsewhere.



Figure 3.

and the last reductive elimination steps are the ratedetermining steps with relative rate r_R/r_S being 49. In the case of the (Z)-enone 7 it probably reacts after being isomerized to the (E)-enone through the routes shown in Scheme II.¹⁹ Addition of TMSCl accelerates the steps following the d- π^* complex (C) formation and drives the reaction all the way to the silyl enol ether (8 or 10). Thus the reaction is kinetically controlled and the stereoselectivity is assumed to be determined by the stereochemistry of the starting enones. The (E)-enone 6 adopts two equally stable eclipsed conformations (A and B, Figure 3) by a molecular mechanics calculation.¹³ Applying Felkin-Anh concept,²⁰ we assumed that the cuprate approaches the conformation A' as shown in Figure 3 to yield d- π^* complex C_1 . In the case of the (Z)-enone 7, B is the only global minimum energy conformation and the reaction is assumed to proceed via a transition state B' to give complex C_2 .

The carbonyl oxygen at C(24) of 9 and 11 was removed via methyl thiocarbonate²¹ of the 24-alcohols (12 and 14) to give desired side chain structure. Other methods, such as reduction of tosylhydrazone,²² reduction of 24-alcohol tosylate (mesylate), and radical deoxygenation via 24phenyl thiocarbonate,^{21b} gave unsatisfactory results. The 24-ketones were reduced (NaBH₄), the resulting epimeric 24-alcohols 12 and 14 were converted to methyl thiocarbonates 13 and 15 (imidazole, NaH, CS₂, and then MeI), and then they were reductively deoxygenated (Bu₃SnH, AIBN) to give 16 and 17 in 74 and 50% yields from 9 and 11, respectively. The hydroxyl protecting groups at C(1), C(3), and C(25) were removed in two steps to give provitamin D (20 and 21) in 80-90% overall yield from 16 and 17. The provitamin D (20 and 21) were converted to (22R)- and (22S)-22-methyl- 1α ,25-dihydroxyvitamin D₃ (1b and 1c) by photochemical reaction followed by thermal isomerization.

The ability of these two 22-methylvitamin D analogs (1b and 1c) to bind to pig intestinal receptor was examined in compared with 1,25-(OH)₂D₃ (1a). The 22S-epimer 1c is about a third as active as the active vitamin D (1a) but the 22R-epimer (1b) was considerably less active (1/50) than la. The results suggest that the side chain conformation required to bind to the receptor is the anti at C(17-20-22-23). Details of the biological activities and the structure-activity studies will be reported elsewhere.

We designed and synthesized active vitamin D analogs (1b and 1c) having a side chain with restricted conformational flexibility. The syntheses used highly diastereoselective conjugate addition of Me₂CuLi to steroidal (E)- and (Z)-22-en-24-ones (6 and 7) as the key step. We suggested a mechanism for the stereoselective cuprate addition. Of the two analogs (1b and 1c) synthesized only the 22S-isomer 1c showed high affinity for VDR indicating that the anti conformer with respect to the C(17-20-22-23) bond is responsible for binding to VDR. This conclusion is in accord with that obtained with 22-methoxy analogs.9

Experimental Section

General. ¹H NMR spectra were measured at 270 MHz on a commercially available instrument. Low and high resolution mass spectra were measured at 70 eV. Relative intensities are given in parentheses. All air-sensitive reactions were run under argon atmosphere, and reagents were added through septa using ovendried syringes. The phrase "dried and evaporated" indicates drying with Na₂SO₄, followed by evaporation of the solvents under house vacuum.

1α,3β-Bis[(methoxycarbonyl)oxy]-23,24-dinor-5,7-choladien-22-al (3). Dimethyl sulfoxide (3.67 mL, 51.8 mmol) in CH₂-Cl₂ (10 mL) was added to a solution of oxalyl chloride (2.07 mL, 23.7 mmol) in CH₂Cl₂ (50 mL) at -78 °C. The mixture was stirred at -78 °C for 15 min, and then a solution of alcohol 2 (10 g, 21.6 mmol) in CH_2Cl_2 (40 mL) was added. After 15 min, triethylamine (15 mL, 108 mmol) was added at that temperature, and then the reaction mixture was allowed to warm to room temperature. The mixture was poured into ice-water and extracted with CH₂Cl₂. The extract was washed with water, dried, and evaporated to give a crystalline residue. The residue was recrystallized from ether- CH_2Cl_2 to give aldehyde 3 (8.18 g, 82%) as colorless needles. The filtrates were evaporated and chromatographed on silica gel (30 g) with CHCl₃ to give 3 (1.40 g, 14%): mp 171-173 °C; ¹H NMR δ 0.66 (3 H, s, H-18), 1.02 (3 H, s, H-19), 1.14 (3 H, d, J = 6.9 Hz, H-21), 3.78 and 3.79 (each 3 H, s, CH₃O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.40 and 5.69 (each 1 H, m, H-6 and -7), 9.59 (1 H, d, J = 3.0 Hz, H-22); IR (KBr) 2962, 2878, 1746, 1723, 1441, 1280, 1257, 1147, 990, 963, 940 cm⁻¹; MS m/z 384 (M⁺ -CH₃OCO₂H, 15), 308 (99), 293 (14), 251 (17), 235 (45), 209 (22), 197 (24), 155 (35), 141 (81), 59 (100). Anal. Calcd for C₂₆H₃₆O₇·1/4H₂O: C, 67.15; H, 7.91. Found: C, 67.25; H, 7.85.

1α,3β-Bis[(methoxycarbonyl)oxy]-22-hydroxy-25-(tetrahydropyranyloxy)-5,7-cholestadien-24-one (4). To a solution of diisopropylamine (1.84 mL, 13.0 mmol) in dry THF (6 mL) was added dropwise n-BuLi (1.6 M in hexane, 6.54 mL, 10.5 mmol) at -78 °C and the solution was stirred at that temperature. After 20 min, to the LDA solution was added dropwise a solution of 3-hydroxy-3-methyl-2-butanone tetrahydropyranyl ether (2.42 g, 13.0 mmol) in dry THF (3 mL) at -78 °C and the mixture was stirred for 15 min. The solution of the enolate was then added to a solution of aldehyde 3 (3.0 g, 6.52 mmol) in dry THF (6 mL) via a double-headed needle at -78 °C, and the mixture was stirred for 90 min at that temperature. The mixture was quenched with aqueous NH₄Cl and extracted with AcOEt. The organic layer was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (70g) with 5-15% AcOEt-benzene to give the coupling product 4 (4.01 g, 95.2%) as a mixture of 22-epimers. 4: ¹H NMR δ 3.78 and 3.79 (each 3 H, s, CH₃O), 4.60 (1 H, m, acetal H), 4.85 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38

⁽¹⁹⁾ If this mechanism is correct, isomerization of the starting enone 7 to (E)-enone 6 must be observed in the thermodynamically controlled cuprate addition reactions. In the conjugate additions of organocuprates to enones, isomerization of (Z)-enone to (E)-enone has frequently been observed.^{12a,17e} In the reaction of (Z)-enone 7 with Me₂CuLi under thermodynamically controlled conditions, however, no Z-E isomerization was detected: When the reaction of the (Z)-enone 7 with Me₂CuLi under thermodynamic conditions was terminated at the middle of the reaction, the recovered enone contained no E-isomer 6. The results can be explained as follows: isomerization of the (Z)-enone 7 to the E-isomer 6 did occur under the reaction conditions but since the E-isomer 6 is far more reactive than the Z-isomer 7 no E-enone remained in the reaction mixture. This assumption was substantiated by a competition experiment. When a mixture of the (E)- and (Z)-enones (6 and 7, 1:1) was treated with 1 equiv of Me₂CuLi (THF, -10 °C), all the *E*-isomer was consumed within 5 min but only 20% of the Z-isomer reacted even after 40 min. These results suggest that in the case of the reaction of the Z-isomer 7, a considerably high energy barrier exists between the d- π^* complexes (C) and the Cu-(III)- β -adducts (D). This means that the rate determining step of the reaction of the (Z)-enone is the isomerization to the (E)-enone thus supporting the proposed mechanism.^{18b} (20) Cherest, M.; Felkin, H.; Prudent, N. Tetrahedron Lett. **1968**, 2199

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and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 3496, 2962, 1748, 1444, 1286, 1151, 1129, 1077, 1023, 984 cm⁻¹; MS m/z 410 (M⁺ – CH₃OCO₂H × 2 – dihydropyran, 0.6), 392 (0.8), 384 (3), 343 (1), 308 (25), 141 (22), 85 (100); HRMS m/z calcd for C₃₆H₅₄O₁₀ 646.3713 (M⁺), found 646.3701.

 $(22E)-1\alpha,3\beta$ -Bis[(methoxycarbonyl)oxy]-25-hydroxy-5,7,-22-cholestatrien-24-one (5). To a solution of β -hydroxy ketone 3 (4.0 g, 6.2 mmol) in toluene-CHCl₃ (3:1, 80 mL) were added p-toluenesulfonic acid monohydrate (115 mg, 0.60 mmol) and magnesium sulfate (2.6 g, 21.6 mmol) and the solution was stirred at 50 °C for 1 h. The reaction mixture was cooled to room temperature, diluted with AcOEt, and washed with 5% NaHCO₃ and water. The organic layer was dried and evaporated. The residue was chromatographed on silica gel (100 g, 5-10% AcOEtbenzene) to afford (E)-enone 5 (2.86 g, 85%): mp 177-178.5 °C (benzene-hexane); ¹H NMR δ 0.66 (3 H, s, H-18), 1.02 (3 H, s, H-19), 1.13 (3 H, d, J = 6.4 Hz, H-21), 1.38 (6 H, s, H-26 and -27), 3.78 and 3.79 (each 3 H, s, CH₃O), 3.98 (1 H, s, OH), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7), 6.35 (1 H, d, J = 15.3 Hz, H-23), 7.00 (1 H, dd, J = 15.3 and 8.9 Hz, H-22); IR (KBr) 3440, 2962, 1744, 1698, 1626, 1444, 1282, 1147, 1067, 994, 791 cm⁻¹; MS m/z 468 (M⁺ – CH₃OCO₂H, 6), 392 (17), 349 (17), 334 (9), 59 (100); HRMS m/z calcd for $C_{31}H_{44}O_8$ 544.3033 (M⁺), found 544.3058. Anal. Calcd for C₃₁H₄₄O₈·1/ 2H₂O: C, 67.25; H, 8.19. Found: C, 67.66; H, 8.17.

 $(22E)-1\alpha, 3\beta$ -Bis[(methoxycarbonyl)oxy]-25-[(methoxymethyl)oxy]-5,7,22-cholestatrien-24-one (6). To a solution of (E)-enone 5 (2.24 g, 4.12 mmol) in dry CH_2Cl_2 (10 mL) were added N.N-diisopropylethylamine (21.5 mL, 123 mmol) and chloromethyl methyl ether (6.25 mL, 82.3 mmol), and the mixture was stirred at ambient temperature. After 3 h, water was added and the mixture was extracted with AcOEt. The extracts were washed with 5% NaHCO3 and water, dried, and evaporated. The residue was chromatographed on silica gel (50 g, 5% AcOEtbenzene) to afford 6 (2.18 g, 90%): mp 152-155 °C (AcOEtbenzene); ¹H NMR δ 0.65 (3 H, s, H-18), 1.02 (3 H, s, H-19), 1.12 (3 H, d, J = 6.9 Hz, H-21), 1.38 (6 H, s, H-26 and -27), 3.38 (3 H)H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO), 4.66 (2 H, s, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7), 6.61 (1 H, d, J = 15.3 Hz, H-23), 6.88 (1 H, dd, J = 15.3, 8.9 Hz, H-22); IR (KBr) 3422, 2956, 1742, 1700, 1632, 1446, 1286, 1257, 1147, 1040 cm⁻¹; UV (95% EtOH) λ_{max} 235, 261, 271, 281.5, 293.5 nm, λ_{min} 255.5, 264.5, 276, 289 nm; MS m/z 512 (M+ - CH₃OCO₂H, 4), 437 (4), 378 (18), 333 (5), 155 (18), 103 (100). Anal. Calcd for C₃₃H₄₈O₉: C, 67.32; H, 8.22. Found: C, 67.44; H, 8.28.

(22Z)-1a,3\beta-Bis[(methoxycarbonyl)oxy]-25-[(methoxymethyl)oxy]-5,7,22-cholestatrien-24-one (7). A solution of (E)enone 6 (180 mg, 0.306 mmol) and naphthalene (100 mg, 0.781 mmol) in benzene (33 mL) in Pyrex vessel was flushed with Ar for 15 min and then irradiated externally with a 100-W highpressure mercury lamp (Shigemi Standard, Tokyo) at room temperature under Ar, until approximately half the (E)-enone was consumed (analyzed by HPLC). The solvent was evaporated and the residue was chromatographed on silica gel (30 g, 3%)AcOEt-benzene) to give (Z)-enone 7 (61 mg, 34%), starting (E)enone 6 (85 mg, 47%), and 20-alcohol 22 (22 mg, 12%) as a 1:1 mixture of 20-epimers, which could not be separated on TLC or HPLC. 7: mp 139-141 °C (MeOH-AcOEt); ¹H NMR δ 0.68 (3 H, s, H-18), 1.02 (3 H, s, H-19), 1.03 (3 H, d, J = 7.4 Hz, H-21), 1.37 (6 H, s, H-26 and -27), 3.38 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO), 4.67 (2 H, s, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.36 and 5.68 (each 1 H, m, H-6 and -7), 6.00 (1 H, dd, J = 11.4, 10.9 Hz, H-22), 6.49 (1 H, d, J = 11.4 Hz, H-23);IR (KBr) 3436, 2958, 1748, 1694, 1620, 1444, 1288, 1257, 1147, 1038 cm⁻¹; UV (95% EtOH) λ_{max} 235, 261, 271, 281.5, 293.5 nm, λ_{\min} 255.5, 264, 276, 289 nm; MS m/z 512 (M⁺ – CH₃OCO₂H, 6), 436 (8), 378 (6), 155 (20), 103 (100), 81 (23). Anal. Calcd for C33H48O9: C, 67.32; H, 8.22. Found: C, 67.36; H, 8.31. 22 (1:1 mixture): ¹H NMR δ 0.65 and 0.69 (3 H (1:1), s, H-18), 0.99 (3 H, s, H-19), 1.386 and 1.391 (6 H (1:1), s, H-26 and -27), 1.50 and 1.52 (3 H (1:1), s, H-21), 3.39 (3 H, s, CH₃OCH₂), 3.77 (3 H, s, CH₃OCO), 3.78 and 3.79 (3 H (1:1), s, CH₃OCO), 4.66 and 4.67 (2 H (1:1), s, OCH₂O), 4.82 (1 H, m, H-1), 4.89 (1 H, m, H-3), 5.36 and 5.68 (each 1 H, m, H-6 and -7), 6.81 and 6.86, (1 H (1:1), d, J = 16 Hz, H-22 or -23), 7.02 and 7.09 (1 H (1:1), d, J = 16 Hz,

H-23 or -22); IR (KBr) 3420, 2962, 2878, 1748, 1698, 1628, 1446, 1367, 1284, 1149, 1083, 1035, 994, 957, 938 cm⁻¹; UV (10% 2-propanol/hexane) λ_{max} 237, 262, 272, 282, 294 nm; MS *m/z* 528 (M⁺ - CH₃OCO₂H, 4), 452 (4), 394 (9), 349 (6), 251 (24), 155 (34), 141 (41), 103 (100); HRMS *m/z* calcd for C₃₃H₄₈O₁₀ 604.3247 (M⁺), found 604.3240.

(22S,23Z)-22-Methyl-5,7,23-cholestatriene-1α,3β,24,-25-tetrol 1,3-Bis(methyl carbonate) 25-(Methoxymethyl) 24-(Trimethylsilyl) Diether (8). Methyllithium (1.4 M in ether, 970 μ L, 1.36 mmol) was added to a suspension of cuprous iodide (130 mg, 0.68 mmol) in THF (400 μ L) at 0 °C and the solution was stirred for 15 min at that temperature. After being cooled to -78 °C, TMSCl (108 µL, 0.85 mmol), HMPA (148 µL, 0.85 mmol), and a solution of (E)-enone 6 (100 mg, 0.17 mmol) in THF (400 μ L) were added in this order and the mixture was stirred at that temperature. After 30 min, triethylamine (2 mL) was added and then the mixture was diluted with AcOEt. The mixture was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10g) with 3% AcOEtbenzene to afford TMS ether 8 (109 mg, 95%): ¹H NMR δ 0.20 (9 H, s, TMS), 0.60 (3 H, s, H-18), 0.88 (3 H, d, J = 6.9 Hz, H-21or 22-Me), 0.94 (3 H, d, J = 6.9 Hz, 22-Me or H-21), 1.00 (3 H, s, H-19), 1.33 and 1.34 (each 3 H, s, H-26 and -27), 3.36 (3 H, s, CH₃OCH₂), 3.77 and 3.79 (each 3 H, s, CH₃OCO), 4.62 and 4.66 $(each 1 H, d, J = 6.9 Hz, OCH_2O), 4.67 (1 H, d, J = 9.4 Hz, H-23),$ 4.83 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.37 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 2964, 1750, 1657, 1444, 1259, 1149, 1098, 1040, 963, 845 cm⁻¹; MS m/z 538 (M⁺ – CH₃OCO₂H – CH₃-OCH₂OH, 15), 462 (6), 372 (3), 354 (5), 185 (41), 183 (44), 141 (35), 131 (31), 111 (100), 83 (25), 73 (98). Anal. Calcd for C₃₇H₆₀O₉Si·1/4H₂O: C, 65.21; H, 8.95. Found: C, 65.17; H, 9.10.

(22R)-1α,3β-Bis[(methoxycarbonyl)oxy]-25-[(methoxymethyl)oxy]-22-methyl-5,7-cholestadien-24-one (9). Deprotection of Enol Silyl Ether 8. To a solution of TMS ether 8 (100 mg, 0.15 mmol) in dry THF (0.8 mL) was added n-Bu₄NF (1.0 M in THF, 150 μ L, 0.15 mmol) and the mixture was stirred at room temperature. After 1.5 h, the reaction was quenched with saturated NH4Cl and extracted with AcOEt. The organic layer was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10g) with 4% AcOEt-benzene to afford 9 (84 mg, 94%): mp 162-165 °C (hexane-AcOEt); 1H NMR δ 0.63 (3 H, s, H-18), 0.853 (3 H, d, J = 5.9 Hz, H-21 or 22-Me), 0.855 (3 H, d, J = 6.9 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.33 and 1.36 (each 3 H, s, H-26 and -27), 3.39 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO₂), 4.72 (2 H, s, OCH₂O), 4.85 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.69 (each 1 H, m, H-6 and -7); IR (KBr) 2962, 1746, 1717, 1444, 1377, 1286, 1149, 1035, 992, 963, 936, 868, 791 cm⁻¹; MS m/z 572 (M⁺ - MeOH, 1), 528 (1), 496 (2), 483 (3), 452 (5), 420 (4), 407 (4), 394 (7), 141 (24), 103 (100). Anal. Calcd for C₃₄H₅₂O₉: C, 67.52; H, 8.67. Found: C, 67.47; H, 8.70.

Addition of Me₂CuLi to (*E*)-Enone 6. Methyllithium (1.4 M in ether, 2.91 mL, 4.07 mmol) was added to a suspension of CuI (389 mg, 2.04 mmol) in dry THF (0.6 mL) at 0 °C and the solution was stirred for 15 min. (*E*)-Enone 6 (300 mg, 0.51 mmol) in THF (1.8 mL) was added to the resulting solution of Me₂CuLi and the mixture was stirred at 0 °C for 25 min. The reaction was quenched with saturated NH₄Cl and extracted with AcOEt. The organic extracts were washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10 g, 5% AcOEtbenzene) to afford 9 together with a small amount of its 22-epimer 11 (98:2 by HPLC analysis, 281 mg, 91%).

Addition of Me₂CuLi to (Z)-Enone 7. (Z)-Enone 7 (100 mg, 0.17 mmol) was treated with Me₂CuLi under similar conditions described above for (E)-enone 6 to yield 9 together with a trace of its 22-epimer 11 (98:2, 80 mg, 78%).

(22*R*,23*Z*)-22-Methyl-5,7,23-cholestatriene- 1α ,3 β ,24,-25-tetrol 1,3-Bis(methyl carbonate) 25-(Methoxymethyl) 24-(Trimethylsilyl) Diether (10). Methyllithium (1.4 M in ether, 2.14 mL, 3.00 mmol) was added to a suspension of cuprous iodide (285 mg, 1.50 mmol) in THF (400 μ L) at 0 °C and the solution was stirred for 15 min at that temperature. The mixture was cooled to -78 °C, TMSCl (237 μ L, 1.87 mmol), HMPA (325 μ L, 1.87 mmol), and a solution of (*Z*)-enone 7 (110 mg, 0.187 mmol) in THF (400 μ L) were added in this order, and the mixture was stirred for 40 min at that temperature. Triethylamine (2 mL) was added at -78 °C and then the mixture was diluted with AcOEt. The mixture was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (12 g, 2% AcOEt-benzene) to afford byproduct 23 (7 mg, 6%) and TMS ether 10 containing a trace of its 22-epimer 8 (97:3 by HPLC, 95 mg, 75%) in this order. 10: ¹H NMR δ 0.20 (9 H, s, TMS) 0.61 (3 H. s. H-18), 0.79 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.86 (3 H, d, J = 5.9 Hz, 22 -Me or H-21), 1.01 (3 H, s, H-19), 1.35 (6)H, s, H-26 and -27), 3.36 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO₂), 4.63 and 4.66 (each 1 H, d, J = 7.4 Hz, OCH₂O), 4.84 (1 H, m, H-1), 4.85 (1 H, d, J = 9.4 Hz, H-23), 4.90 (1 H, m, H-23)H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 2964, 1748, 1657, 1446, 1282, 1149, 1042, 963, 843 cm⁻¹; MS m/z 538 $(M^+ - CH_3OCO_2H - CH_3OCH_2OH, 1), 462 (1), 390 (1), 372 (1),$ 183 (100), 141 (9), 111 (11), 73 (74); HRMS m/z calcd for C₃₇H₆₀O₉-Si 676.4002 (M⁺), found 676.3962. 23: ¹H NMR & 0.20 (9 H, s, TMS), 0.65 (3 H, s, H-18), 1.02 (3 H, s, H-19), 1.07 (3 H, d, J = 6.4 Hz, H-21), 1.68 and 1.72 (each 3 H, s, H-26 and -27), 3.77 and 3.79 (each 3 H, s, CH₃OCO), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.37 and 5.68 (each 1 H, m, H-6 and -7), 5.59 (1 H, dd, J = 15.3 and 8.9 Hz, H-22), 6.20 (1 H, d, J = 15.3 Hz, H-23); IR (KBr) 2960, 2876, 1748, 1626, 1446, 1282, 1199, 1149, 984, 963, 940 cm⁻¹; UV (95% EtOH) λ_{max} 251, 260, 270.5, 281.5, 293.5 nm, λ_{\min} 221.5, 267, 277, 289 nm; MS m/z 524 (M⁺ – CH₃OCO₂H, 5), 448 (12), 433 (2), 376 (2), 251 (8), 249 (13), 197 (36), 170 (30), 73 (100); HRMS m/z calcd for C₃₄H₅₂O₇Si 600.3483 (M⁺), found 600.3477.

(22.S)- 1α , 3β -Bis[(methoxycarbonyl)oxy]-25-[(methoxymethyl)oxy]-22-methyl-5,7-cholestadien-24-one (11). Deprotection of Enol Silyl Ether 10. TMS ether 10 (160 mg, 0.24 mmol) was deprotected similarly to give 11 (131 mg, 92%): ¹H NMR δ 0.60 (3 H, s, H-18), 0.73 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.82 (3 H, d, J = 5.4 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.34 (6 H, s, H-26 and -27), 3.39 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO₂), 4.71 (2 H, s, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.69 (each 1 H, m, H-6 and -7); IR (KBr) 2966, 1746, 1717, 1444, 1284, 1149, 1033, 938, 793 cm⁻¹; MS m/z 572 (M⁺ – CH₃OH, 1), 528 (3), 497 (1), 452 (7), 394 (8), 141 (19), 103 (100); HRMS m/z calcd for C₃₄H₅₂O₉ 604.3608 (M⁺), found 604.3611.

(22R,24R)- and (22R,24S)-22-Methyl-5,7-cholestadiene- $1\alpha, 3\beta, 24, 25$ -tetrol 1,3-Bis(methyl carbonate) 25-(Methoxymethyl) Ether (12a and 12b). To a solution of 9 (213 mg, 0.35 mmol) in CH₂Cl₂ (2 mL) at room temperature were added methanol (1 mL) and NaBH₄ (27 mg, 0.71 mmol) and the mixture was stirred for 70 min. The solution was diluted with CH_2Cl_2 and water and extracted with CH₂Cl₂. The organic layer was washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10g, 5% AcOEt-benzene) to afford 12a (158 mg, 74%) and its 24S-epimer 12b (52 mg, 24%) in this order. 12a: mp 156-157.5 °C (MeOH); 1H NMR & 0.63 (3 H, s, H-18), 0.81 (3 H, d, J = 5.4 Hz, H-21 or 22-Me), 0.91 (3 H, d, J = 6.4 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.17 and 1.20 (each 3 H, s, H-26 and -27), 3.40 (3 H, s, CH₃OCH₂), 3.42 (1 H, m, H-24), 3.77 and 3.79 (each 3 H, s, CH₃OCO₂), 4.71 and 4.78 (each 1 H, d, J = 7.4 Hz, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.69 (each 1 H, m, H-6 and -7); IR (KBr) 3466, 2960, 2878, 1748, 1444, 1373, 1270, 1147, 1038 cm⁻¹; MS m/z 530 (M⁺ - CH₃OCO₂H, 8), 498 (5), 454 (16), 422 (48), 392 (41), 351 (38), 251 (34), 224 (24), 223 (24), 209 (72), 197 (39), 155 (59), 141 (60), 103 (100), 72 (91), 59 (60), 55 (73). Anal. Calcd for C₃₄H₅₄O₉: C, 67.30; H, 8.97. Found: C, 67.48; H, 9.15. 12b: 1H NMR & 0.62 (3 H, s, H-18), 0.84 (3 H, d, J = 5.4 Hz, H-21 or 22-Me), 1.01 (3 Hz)H, s, H-19), 1.04 (3 H, d, J = 6.9 Hz, 22-Me or H-21), 1.17 and 1.21 (each 3 H, s, H-26 and -27), 3.39 (3 H, s, CH₃OCH₂), 3.42 (1 H, m, H-24), 3.77 and 3.79 (each 3 H, s, CH₃OCO₂), 4.70 and 4.77 $(each 1 H, d, J = 7.4 Hz, OCH_2O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-1), 4.90 (1 H, m, H-1))$ m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 3448, 2962, 1746, 1444, 1274, 1147, 1036 cm⁻¹; MS m/z 530 (M⁺ – CH₃-OCO₂H, 3), 498 (6), 454 (8), 422 (29), 392 (48), 351 (33), 251 (32), 224 (20), 223 (21), 209 (55), 197 (34), 155 (53), 141 (50), 103 (100), 72 (82), 59 (53), 55 (65); HRMS m/z calcd for C₃₄H₅₄O₉ 606.3767 (M⁺), found 606.3763.

(22S)-22-Methyl-5,7-cholestadiene-1α,3β,24,25-tetrol 1,3-Bis(methyl carbonate) 25-(Methoxymethyl) Ether (14). 22-Methyl ketone 11 (91 mg, 0.15 mmol) was similarly reduced with NaBH₄ to give 24-alcohol 14 (84 mg, 92%) as a mixture of 24epimers (1:1 by NMR). 14: ¹H NMR δ 0.62 and 0.64 (3 H (1:1), s, H-18), 1.01 and 1.02 (3 H (1:1), s, H-19), 3.39 (3 H, s, CH₃-OCH₂), 3.77 and 3.79 (each 3 H, s, CH₃OCO₂), 4.72 and 4.76 (each 1 H, d, J = 7.4 Hz, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 3464, 2964, 1744, 1446, 1255, 1147, 1033, 955, 795 cm⁻¹; MS m/z 530 (M⁺ – CH₃OCO₂H, 6), 498 (5), 454 (12), 422 (51), 392 (29), 351 (32), 251 (44), 224 (35), 223 (27), 209 (87), 197 (64), 155 (72), 141 (76), 103 (100), 72 (99), 59 (52), 55 (66); HRMS m/z calcd for C₃₄H₅₄O₉ 606.3767 (M⁺), found 606.3751.

(22R, 24R)-22-Methyl-5,7-cholestadiene-1 α , 3 β , 24,-25-tetrol 1.3-Bis(methyl carbonate) 24-(S-Methyl dithiocarbonate) 25-(Methoxymethyl) Ether (13a). To a solution of 24-alcohol 12a (100 mg, 0.16 mmol) and imidazole (0.2 mg) in THF (1.5 mL) at room temperature was slowly added NaH (50% oil dispersion, 12 mg, 0.25 mmol). After the mixture was stirred for 20 min, carbon disulfide (30 μ L, 0.50 mmol) was added all at once. Stirring was continued for 30 min, and then iodomethane $(20 \ \mu L, 0.32 \ mmol)$ was added in a single portion. The mixture was stirred another 30 min and quenched with water. The mixture was extracted with AcOEt, and the extracts were washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10 g, 2% AcOEt-benzene) to afford 13a (102 mg, 89%): ¹H NMR δ 0.60 (3 H, s, H-18), 0.83 (3 H, d, J = 5.4 Hz, H-21 or 22-Me), 0.98 (3 H, d, J = 6.4 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.27 (6 H, s, H-26 and -27), 2.58 (3 H, s, SCH₃), 3.37 (3 H, s, CH₃OCH₂), 3.77 and 3.79 (each 3 H, s, CH₃OCO₂), 4.70 and 4.79 (each 1 H, d, J = 7.4 Hz, OCH₂O), 4.85 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.37 and 5.68 (each 1 H, m, H-6 and -7), 5.90 (1 H, d, J = 10.4 Hz, H-24); MS m/z 467 (M⁺ – CH₃OCO₂H × 2 - CH₃OCH₂OH - CH₃, 38), 392 (36), 325 (31), 251 (42), 235 (21), 223 (29), 209 (76), 197 (50), 183 (28), 155 (83), 141 (68), 123 (31), 113 (34), 91 (89), 69 (100), 55 (81).

(22*R*,24*S*)-22-Methyl-5,7-cholestadiene- 1α ,3 β ,24,-25-tetrol 1,3-Bis(methyl carbonate) 24-(*S*-Methyl dithiocarbonate) 25-(Methoxymethyl) Ether (13b). The epimeric alcohol 12b was converted to *S*-methyl dithiocarbonate 13b under similar conditions. 13b: ¹H NMR δ 0.61 (3 H, s, H-18), 0.83 (3 H, d, *J* = 5.9 Hz, H-21 or 22-Me), 0.93 (3 H, d, *J* = 6.9 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.27 (6 H, s, H-26 and -27), 2.58 (3 H, s, SCH₃), 3.37 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO₂), 4.71 and 4.77 (each 1 H, d, *J* = 7.4 Hz, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.69 (each 1 H, m, H-6 and -7), 5.88 (1 H, dd, *J* = 8.9, 3.0 Hz, H-24).

(22.S)-22-Methyl-5,7-cholestadiene- $1\alpha_3\beta_2$,24,25-tetrol 1,3-Bis(methyl carbonate) 24-(S-Methyl dithiocarbonate) 25-(Methoxymethyl) Ether (15). (22S)-24-Alcohol 14 (26 mg, 0.043 mmol) was converted to dithiocarbonate 15 (18 mg, 60%) under similar conditions. 15: ¹H NMR δ 0.60 and 0.62 (3 H (1:1), s, H-18), 1.01 (3 H, s, H-19), 1.27 (6 H, s, H-26 and -27), 2.565 and 2.569 (3 H (1:1), s, SCH₃), 3.367 and 3.370 (3 H (1:1), s, CH₃-OCH₂), 3.775 and 3.782 (each 3 H, s, CH₃OCO₂), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.36 and 5.68 (each 1 H, m, H-6 and -7), 5.91 (1 H, m, H-24); IR (KBr) 2964, 1748, 1267, 1058 cm⁻¹; MS m/z 512 (M⁺ – CH₃OCO₂H × 2 – CH₃OH, 4), 467 (3), 450 (5), 392 (20), 374 (18), 359 (12), 325 (7), 277 (22), 251 (32), 249 (35), 223 (31), 209 (75), 197 (55), 183 (28), 155 (64), 141 (64), 123 (29), 95 (50), 69 (63), 60 (100), 55 (75); HRMS m/z calcd for C₃₆H₅₆O₉S₂ 696.3366 (M⁺), found 696.3358.

(22*R*)-22-Methyl-5,7-cholestadiene- 1α ,3 β ,25-triol 1,3-Bis-(methyl carbonate) 25-(Methoxymethyl) Ether (16). To a solution of dithiocarbonate 13a (140 mg, 0.20 mmol) in dry toluene (2.5 mL) were added azobis(isobutyronitrile) (8 mg, 0.05 mmol) and *n*-Bu₃SnH (161 μ L, 0.60 mmol), and the mixture was refluxed for 15 min. The reaction mixture was directly chromatographed on silica gel (7 g, 15% AcOEt-hexane) to afford 16 (101 mg, 85%): ¹H NMR δ 0.61 (3 H, s, H-18), 0.82 (3 H, d, J = 6.4 Hz, H-21 or 22-Me), 0.88 (3 H, d, J = 6.9 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.21 (6 H, s, H-26 and -27), 3.37 (3 H, s, CH₃OCH₂), 3.77 and 3.79 (each 3 H, s, CH₃OCO₂), 4.70 (2 H, s, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.37 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 2958, 1744, 1444, 1255, 1147, 1044, 955, 868, 793 cm⁻¹; MS m/z 452 (M⁺ - CH₃OCO₂H - CH₃OCH₂OH, 14), 438 (7), 376 (100), 361 (12), 325 (14), 277 (25), 249 (25), 209 (55), 197 (27), 155 (39), 141 (41), 103 (43), 69 (50), 55 (57); HRMS m/z calcd for $C_{34}H_{54}O_8$ 590.3819 (M⁺), found 509.3809.

(22S)-22-Methyl-5,7-cholestadiene-1α,3β,25-triol 1,3-Bis-(methyl carbonate) 25-(Methoxymethyl) Ether (17). Under similar conditions, dithiocarbonate 15 (34 mg, 0.049 mmol) was reduced to give 17 (26 mg, 90%): ¹H δ 0.62 (3 H, s, H-18), 0.72 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.79 (3 H, d, J = 5.9 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.21 (6 H, s, H-26 and -27), 3.37 (3 H, s, CH₃OCH₂), 3.78 and 3.79 (each 3 H, s, CH₃OCO₂), 4.71 (2 H, s, OCH₂O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7); IR (KBr) 2962, 1744, 1444, 1284, 1147, 1042, 955, 866, 793 cm⁻¹; MS m/z 452 (M⁺ - CH₃-OCO₂H - CH₃OCH₂OH, 11), 438 (8), 376 (100), 361 (16), 277 (28), 251 (32), 249 (29), 224 (31), 209 (78), 197 (49), 155 (51), 141 (59), 103 (42), 69 (81), 55 (79); HRMS m/z calcd for C₃₄H₅₄O₈ 590.3818 (M⁺), found 590.3826.

(22R)-22-Methyl-5,7-cholestadiene-1a,36,25-triol1,3-Bis-(methyl carbonate) (18). A solution of MOM ether 16 (101 mg, 0.17 mmol) and p-toluenesulfonic acid monohydrate (90 mg, 0.47 mmol) in 95% ethanol (4 mL) was refluxed for 10 min. The solvent was evaporated, the residue was dissolved in AcOEt, washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10 g, 3% AcOEt-benzene) to afford 25-alcohol 18 (86 mg, 92%): ¹H NMR δ 0.61 (3 H, s, H-18), 0.82 (3 H, d, J = 6.4 Hz, H-21 or 22-Me), 0.88 (3 H, d, J = 6.9 Hz,22-Me or H-21), 1.01 (3 H, s, H-19), 1.210 and 1.216 (each 3 H, s, H-26 and -27), 3.78 and 3.79 (each 3 H, s, CH₃O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.69 (each 1 H, m, H-6 and -7); IR (KBr) 3552, 2966, 1744, 1731, 1446, 1344, 1292, 1265, 1098, 1025, 797 cm⁻¹; MS m/z 470 (M⁺ – CH₃OCO₂H, 8), 394 (81), 379 (17), 376 (19), 361 (6), 251 (18), 224 (26), 209 (67), 197 (39), 155 (49), 141 (58), 69 (69), 59 (100), 55 (75).

(22S)-22-Methyl-5,7-cholestadiene- 1α ,3 β ,25-triol 1,3-Bis-(methyl carbonate) (19). Under similar conditions, MOM ether 17 (11 mg, 0.019 mmol) was converted to 25-alcohol 19 (8 mg, 80%): ¹H NMR δ 0.62 (3 H, s, H-18), 0.73 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.79 (3 H, d, J = 5.9 Hz, 22-Me or H-21), 1.01 (3 H, s, H-19), 1.21 (6 H, s, H-26 and -27), 3.78 and 3.79 (each 3 H, s, CH₃O), 4.84 (1 H, m, H-1), 4.90 (1 H, m, H-3), 5.38 and 5.68 (each 1 H, m, H-6 and -7).

(22*R*)-22-Methyl-5,7-cholestadiene- 1α ,3 β -25-triol (20). To a solution of biscarbonate 18 (80 mg, 0.15 mmol) in CH₂Cl₂ (1 mL) was added 5% KOH/MeOH (3.5 mL) and the mixture was stirred at 40 °C for 45 min. The mixture was concentrated in vacuo and diluted with CHCl₃, washed with water, dried, and evaporated. The residue was chromatographed on silica gel (10 g, 60% AcOEt-CH₂Cl₂) to afford provitamin D 20 (60 mg, 95%): ¹H NMR δ 0.63 (3 H, s, H-18), 0.84 (3 H, d, J = 6.4 Hz, H-21 or 22-Me), 0.88 (3 H, d, J = 6.9 Hz, 22-Me or H-21), 0.95 (3 H, s, H-19), 1.216 and 1.223 (each 3 H, s, H-26 and -27), 3.78 (1 H, m, H-1), 4.07 (1 H, m, H-3), 5.38 and 5.73 (each 1 H, m, H-6 and -7); IR (KBr) 3426, 2934, 1460, 1383, 1263, 1031, 909, 801 cm⁻¹; UV (95% EtOH) λ_{max} 272, 282.5, 294.5 nm, λ_{min} 235, 277, 290 nm; MS m/z 430 (M⁺, 18), 412 (18), 394 (19), 251 (20), 227 (32), 197 (26), 171 (35), 157 (39), 69 (88), 59 (94), 55 (100); HRMS m/z calcd for C₂₈H₄₆O₃ 430.3447 (M⁺), found 430.3434.

(22S)-22-Methyl-5,7-cholestadiene-1α,3β,25-triol (21). Under similar conditions, biscarbonate 19 (19 mg, 0.036 mmol) was

hydrolyzed to give provitamin D 21 (14 mg, 93%): ¹H NMR δ 0.64 (3 H, s, H-18), 0.74 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.81 (3 H, d, J = 5.9 Hz, 22-Me or H-21), 0.95 (3 H, s, H-19), 1.22 (6 H, s, H-26 and -27), 3.77 (1 H, m, H-1), 4.07 (1 H, m, H-3), 5.38 and 5.74 (each 1 H, m, H-6 and -7); IR (KBr) 3408, 2966, 1460, 1381, 1154, 1054, 909, 760 cm⁻¹; UV (95% EtOH) λ_{max} 272, 282.5, 294.5 nm, λ_{min} 233, 277, 290 nm; MS m/z 430 (M⁺, 18), 412 (12), 394 (17), 251 (19), 227 (31), 197 (26), 171 (35), 157 (42), 69 (82), 59 (100), 55 (89); HRMS m/z calcd for C₂₈H₄₆O₃ 430.3447 (M⁺), found 430.3442.

(22R)-22-Methyl-1 α ,25-dihydroxyvitamin D₃ (1b). A solution of the provitamin D 20 (60 mg, 0.14 mmol) in benzene-EtOH (150:20, 170 mL) was flushed with Ar for 15 min and then irradiated at 0 °C under Ar with a 100-W high-pressure mercury lamp (Shigemi Standard, Tokyo) through a Vycor filter until most of the provitamin D was consumed (the reaction was monitored by HPLC). The solvent was evaporated and the residue was chromatographed on Sephadex LH-20 (20 g) with $CHCl_3$ /hexane/MeOH (70:30:1) to give the previtamin D (30.6 mg, yield 51%). The previtamin D was dissolved in 95% ethanol (5 mL) and stored in the dark at room temperature under Ar for 2 weeks. The solvent was evaporated and the residue was chromatographed on Sephadex LH-20 (12g) with the same solvent system as above to give 1b (24.2 mg, 79%) as a colorless glass: ¹H NMR δ 0.54 (3 H, s, H-18), 0.82 and 0.88 (each 3 H, d, J = 6.9 Hz, H-21 and 22-Me), 1.218 and 1.225 (each 3 H, s, H-26 and -27), 4.23 (1 H, m, H-3), 4.43 (1 H, m, H-1), 5.01 and 5.33 (each 1 H, m, H-19), 6.02 and 6.38 (each 1 H, d, J = 11.4 Hz, H-7 and -6, respectively); IR (KBr) 3398, 2952, 2878, 1657, 1543, 1460, 1381, 1261, 1060, 959, 911, 801 cm⁻¹; UV (95% EtOH) λ_{max} 264 nm, λ_{\min} 228 nm; MS m/z 430 (M⁺, 3), 412 (7), 394 (6), 379 (3), 152 (26), 134 (100), 69 (69), 59 (71), 55 (86); HRMS m/z calcd for C₂₈H₄₆O₃ 430.3447 (M⁺), found 430.3436.

(22S)-22-Methyl-1 α ,25-dihydroxyvitamin D₃ (1c). Under similar conditions described above, provitamin D 21 (32.7 mg, 0.076 mmol) was converted to the previtamin D (12.4 mg, yield 38%) and then isomerized to vitamin D (1c) (8.8 mg, 71%): ¹H NMR δ 0.55 (3 H, s, H-18), 0.73 (3 H, d, J = 6.9 Hz, H-21 or 22-Me), 0.78 (3 H, d, J = 5.9 Hz, 22-Me or H-21), 1.21 (6 H, s, H-26 and -27), 4.23 (1 H, m, H-3), 4.43 (1 H, m, H-1), 5.01 and 5.33 (each 1 H, m, H-19), 6.02 and 6.38 (each 1 H, d, J = 10.9 Hz, H-7 and -6, respectively); IR (KBr) 3400, 2940, 1638, 1381, 1156, 1058, 909 cm⁻¹; UV (95% EtOH) λ_{max} 265 nm, λ_{min} 228 nm; MS m/z 430 (M⁺, 3), 412 (6), 394 (5), 379 (3), 152 (24), 134 (100), 69 (60), 59 (67), 55 (71); HRMS m/z calcd for C₂₈H₄₆O₃ 430.3447 (M⁺), found 430.3451.

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Supplementary Material Available: ¹H NMR spectra of compounds 1b, 1c, 4, 10, 11, 12b, 13a, 13b, and 14–23 (18 pages). This material is contained in libraries on microfiche, immediately follows this article in the microfilm version of the journal, and can be ordered from the ACS; see any current masthead page for ordering information.